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Taibaihenryiin C, a diterpenoid with a novel skeleton from Isodon henryi

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Abstract—A diterpenoid with a novel skeleton, taibaihenryiin C, along with four known compounds, odonicin, rabdosianin B, wikestroemioidin C and eupafolin were isolated from an ethanol extract of the leaves and new branches of *Isodon henryi* (Hemsl.) Kudo (Labiatae) and its structure was elucidated on the basis of spectroscopic data and X-ray crystallography. © 2002 Elsevier Science Ltd. All rights reserved.

In past studies, a large number of structurally interesting diterpenoids with various biological activities, such as anti-HIV,¹ antitumor and inhibition of insect growth,² were discovered from medicinal plants of the genus Isodon. Plants of this genus have been used for gastrointestinal disorders and as antitumor and antiphlogistic agents in Chinese traditional medicine.³ Three major structural types, the ent-kaurene type, the 6,7-B-seco-ent-kaurene type and the 8,9-seco-ent-kaurene type and three minor structural types, 6-epi-entgibberelene, ent-isopimanthrene and abietene type have been found in these Isodon diterpenoids.⁴ Recently, our investigations of the chemical constituents of a specimen of Isodon henryi collected on Taibai Mountain, Shaanxi Province, PR China, yielded a diterpenoid with a novel skeleton, taibaihenryiin C 1, together with four known compounds, odonicin 2, rabdosianin B 3, wikestroemioidin C 4 and eupafolin 5.

Taibaihenryiin C 1 was obtained from the dried powdered leaves and new branches extract $(0.0025\%)^5$ as irregular hexagonal colorless crystals (mp 255– 256.5°C), with $[\alpha]_D^{25}$ –111.3 (c=0.55, CHCl₃) and had the molecular formula C₂₂H₃₀O₅ as revealed by the FABMS [m/z: 381(M+Li)⁺ and 397(M+Na)⁺], the Xray single-crystal structure determination, and elemental analysis [found: C, 70.5%; H, 7.8% (calcd C, 70.6%; H, 8.0%)]. Its IR spectrum showed characteristic absorption bands for the hydroxyl group, the carbonyl

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group and double bonds (3537, 1754, 1715, 1666 cm⁻¹), and these assignments were confirmed by its ¹³C and ¹H NMR spectral data (see Table 1) (δ 170.6, 170.8 ppm; 132.7, 130.2 ppm; 142.6, 120.1 ppm and δ 6.08, 6.13, 5.49 ppm). The ¹H, ¹³C and HMQC NMR spectra of **1** showed signals for three methyl groups, four methylene groups, six methine groups, two quaternary carbons and four olefinic carbons including an *exo*-methylene, one acetoxyl group and one ester carbonyl carbon.

Since four out of eight degrees of unsaturation were accounted for, taibaihenryiin C 1 was inferred to contain four rings. In the HMBC spectrum of 1 (see Table 1), the signals at δ 1.59 (H-2 α) and 1.63 (H-2 β) showed correlation with the signals at δ 25.1 (t, C-1), 75.8 (d, C-3) and 37.4 (s, C-4); the signal at δ 1.80 (1H, dd, J=12.1, 4.8 Hz, H-5 β) correlated with the signals at δ 37.4 (s, C-4), 35.4 (s, C-10), 14.3 (q, C-20) and 25.1 (t, C-1); the signals at δ 0.96 (3H, H-18) and 0.92 (3H, H-19) exhibited cross peaks with both signals at δ 37.4 and 43.1 (d, C-5), respectively. These indicated the presence of structure unit ring A with two methyl groups at C-4 and a methyl group at C-10 (see Fig. 1). Ring B was formed by connecting the structural unit -CH₂-CH=C-CH- through C-5 and C-10, which was judged from the HMBC correlation from the two proton signals at δ 2.02 (1H, m, H-6 α) and 2.09 (1H, m, H-6 β) to the carbon signals at δ 43.1 (d, C-5), 132.7 (d, C-7) and 130.2 (s, C-8), and from the proton signal at δ 2.40 (1H, br s, H-9β) to the carbon signals at δ 35.4 (s, C-10), 130.2 (s, C-8), 132.7 (d, C-7), 69.2 (d, C-11) and 14.3 (s, C-20). HMBC correlations from the proton signal at δ 1.71 (1H, dd, J=7.3, 2.2 Hz, H-12α) to the carbon signal at δ 36.9 (d, C-13); from δ 2.03 (1H, m, H-12β) to δ 51.0 (d, C-9), 69.2 (d, C-11) and 81.8 (d, C-14); from δ 3.33 (1H, m, H-13α) to δ 69.2 (d, C-11) and 33.9 (t, C-12), and from δ 4.94 (1H, d, J=8.1 Hz, H-14α) to δ 130.2 (s, C-8), 132.7 (d, C-7), suggested that ring C was constructed by connecting the structural unit -CH-CH₂-CH-CH- through C-9 and C-8. Meanwhile, its ¹³C NMR spectrum showed signals for two ester carbonyl carbons (δ 170.6 and 170.8 ppm),



Figure 1. X-Ray crystal structure of 1.

Table 1. The ¹H, ¹³C NMR, HMBC and ¹H–¹H COSY spectral data of taibaihenryiin C in CD₃Cl^a

Position (H)	$\delta_{\rm H}$, mult., intgr. (J, Hz)	$\delta_{\rm c}$ (mult.) ^b	HMBC (carbon) ^c	¹ H– ¹ H COSY
1α	1.63, m, 1H	25.1 (t)	10, 4	3α
1β	1.90, overlap, 1H		5	3α
2α	1.59, m, 1H	30.6 (t)	(1), (3), 4, 5, 10, 20	n.o. ^d
2β	1.63, overlap, 1H		(1), (3), 4, 5, 10, 20	3α
3α	3.47, t, 1H (2.5)	75.8 (d)	(2), (4), 5, 19	1β, 1α, 2β
4		37.4 (s)		
5β	1.80, dd, 1H (12.1, 4.8)	43.1 (d)	1, (4), 9, (10), 20	n.o.
6α	2.02, m, 1H	23.6 (t)	(5), (7), 8	7
6β	2.09, m, 1H		(5), (7), 8	7
7	6.08, dd, 1H (4.8, 2.2)	132.7 (d)	5, (6), 9, 14	6α, 6β, 9β
8		130.2 (s)		
9β	2.40, br s, 1H	51.0 (d)	(8), 7, (10), (11), 20	7, 11α, 12β
10		35.4 (s)		
11α	5.14, t, 1H (1.6)	69.2 (d)	8, 10, 13, OAc (170.6 ppm)	9β, 12β
12α	1.71, dd, 1H (7.3, 2.2)	33.9 (t)	(13), 16	13α
12β	2.03, m, 1H		9, (11), 14	9β, 11α, 13α
13α	3.33, m, 1H	36.7 (d)	11, (12), 15, 16	12α , 12β , 14α , $17a$, $17b$
14α	4.94, d, 1H (8.1)	81.8 (d)	5, 7, (8), 12	13α
15		170.8 (s)		
16		142.6 (s)		
17a	6.13, d, 1H (2.2)	120.1 (t)	13, 15, (16)	13α
17b	5.49, d, 1H (1.9)		13, 15, (16)	13α
18	0.96, s, 3H	28.2 (q)	(4), 5, 19	n.o.
19	0.92, s, 3H	22.5 (q)	(4), 5, 19	n.o.
20	0.82, s, 3H	14.3 (q)	2, 5, 9, (10)	n.o.
OAc		170.6 (s)		
	1.91, s, 3H	21.0 (q)		

^a The chemical shift values are in parts per million relative to TMS. The spectra were recorded at 500 MHz at room temperature.

^b The ¹³C NMR multiplicities were obtained from the HMQC spectrum.

^c Two-bond correlations are indicated in parentheses.

^d n.o. indicates no clear correlations.

but there was only one signal of the methyl of an acetoxyl group (δ 1.91, s, 3H) in the ¹H NMR spectrum, which revealed that a lactone structure was present in 1. Connecting C-16 and the oxygen atom of the ester group to ring C through C-13 and C-14, respectively, constructed ring D with an exo-methylene γ -lactone structure. This was deduced from the HMBC correlations from the signal at δ 3.33 (1H, m, H-13 α) to δ 142.6 (s, C-16) and 170.8 (s, C-15) and from δ 6.13 (1H, d, J=2.2 Hz, H-17a) and 5.49 (1H, d, J=1.9 Hz, J=1.9 Hz)H-17b) to δ 142.6 (s, C-16), 36.7 (d, C-13) and 170.8 (s, C-15). This evidence indicated that compound 1 was a diterpenoid with a novel skeleton containing a y-lactone. The hydroxyl group and the acetoxyl group in 1 were located at C-3 and C-11 resulting from the signals at δ 3.47 (1H, t, J=2.5 Hz, H-3 α), 75.8 (d, C-3) in the ¹H, ¹³C and HMQC NMR spectra of 1, and the HMBC correlation from the proton signal at δ 5.14 (1H, t, J=1.6 Hz, H-11 α) to the carbon signal at δ 170.6 (s, AcO), respectively. The above conclusions were further confirmed by the ¹H–¹H COSY spectral data of 1 (see Table 1). Thus, the structure of taibaihenryiin C was elucidated to be 1.

The relative stereochemistry of **1** was determined as shown in Fig. 1 by single-crystal X-ray diffraction analysis.⁶ The hydroxyl group at C-3 and the acetoxyl group at C-11 are in the β -orientations, respectively. Ring A is in a chair conformation, and rings B and C approximate in boat conformations. C-14 diverges slightly from the common plane of C-13, C-16, C-15 and O-4, so ring D approximates to an envelope conformation.

Compounds 2, 3 and 4 were identified as the diterpeneoids, odonicin, rabdosianin B and wikestroemioidin C, respectively; compound 5 was shown to be the flavone eupafolin, by comparison of the physical and 13 C NMR data with that reported.⁷

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- 5. The dried powdered leaves and new branches of *Isodon henryi* (5.0 kg) were extracted with 95% EtOH (15 L×3) at room temperature for 7 days. After removal of the solvent in vacuo, the residue was partitioned in H₂O and extracted with petroleum ether (3 L×3) and EtOAc (3 L×3), respectively. The EtOAc extract (122 g) was subjected to column chromatography on silica gel (2 kg, 200–300 mesh), eluting with CHCl₃ and increasing proportions of Me₂CO (CHCl₃/Me₂CO: from 10:0 to 0:10). Fractions were combined by monitoring with TLC. All components were further purified by column chromatography and preparative TLC on silica gel to give 2 (113 mg, 0.0023%), 3 (194 mg, 0.0039%), 1 (126 mg, 0.0025%), 4 (114 mg, 0.0023%) and 5 (623 mg, 0.012%), respectively.
- Crystallographic data (excluding structure factors) have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC 182951. Copies of the data can be obtained free of charge, on application to CCDC, 12 Union Road, Cambridge CB12 1EZ, UK [fax: +44(0)-1223-336033 or e-mail: deposit@ccdc.cam.ac.uk].
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